

High-Dose Statin Flash-Potentiation of Cancer Chemotherapy

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Abstract

In low micromolar concentrations, statins have been shown to boost the apoptotic response to a wide range of cytotoxic agents, in a wide range of cancers. This effect appears to be mediated by inhibition of protein geranylgeranylation, leading to an up-regulation of apoptotic mechanisms. Statin intakes at least an order of magnitude higher than those used standardly for serum lipid modulation may be required to evoke this phenomenon clinically, and such high intakes would evidently entail toxic risk. Nonetheless, several phase I and phase II studies indicate that lovastatin intakes of around 30 mg/kg/day, given for 7 consecutive days once monthly, are reasonably tolerable for most patients. This suggests that it may be feasible to employ high-dose statins as an adjuvant to chemotherapy, if the statins are administered for only several days at a time prior to and following administration of the cytotoxin. Concurrent administration of tocotrienols, which can down-regulate expression of HMG-CoA reductase in cancers, could be expected to amplify the impact of statins on isoprenylation, while likely promoting chemosensitization by independent mechanisms.

Cancer Chemosensitizing Activity of Statins

A number of cell culture studies have demonstrated that, in low micromolar concentrations, statins can dose-dependently potentiate the cell killing efficacy of various cytotoxic agents (e.g. carboplatin, cisplatin, docetaxal, doxorubicin, gemcitabine, 5-fluorouracil, mitomycin C) in a range of cancer cell lines (hepatoma, colon, melanoma, leukemia, breast, pancreatic, non-small cell lung, ovarian, nasopharyngeal, gastric, osteosarcoma).¹⁻²⁰ This phenomenon hence appears to be of some generality, and may reflect a statin-mediated up-regulation of apoptotic mechanisms. In some of these studies, co-administration of geranylgeranyl diphosphate, but not farnesyl diphosphate, abrogates this phenomenon, suggesting that impaired function of G proteins requiring geranylgeranylation, such as those of the Rac/Rho family - and/or other soluble proteins which require post-translational geranylgeranylation for proper integration into cell membranes, such as lamins - mediates the impact of statins in this regard.^{2, 10} A handful of studies in mouse xenograft models indicate that high-dose statin co-administration can boost the cancer-retardant efficacy of cytotoxic agents *in vivo*.^{4, 7, 13, 20} The interaction of statins with doxorubicin is of especial interest, as statins have the potential to decrease the cardiotoxicity of this agent while enhancing its cancer-killing efficacy; this may reflect a key role for Rac1 activity in mediation of this cardiotoxicity.^{13, 21, 22}

How suppressed activity of isoprenylated proteins translates into an up-regulation of chemosensitivity remains unresolved. In statin-treated cancer cell lines, modulated expression of proteins that regulate apoptosis has been described, as well as a reduction in Akt phosphorylation and NF-kappaB activity;^{13, 15, 19, 23-25} why these effects would be downstream consequences of decreased geranylgeranylation requires clarification. Another key consideration that, by and large, is not addressed by the statin chemosensitization literature is whether these effects of statins are relatively specific to transformed cells, as opposed to healthy tissues. However, a report that simvastatin boosts the growth retardant impact of

carmustine in rats bearing gliomas, without however influencing the myelotoxicity of this drug, provides a measure of reassurance.²⁰

An effect of lovastatin independent of its impact HMG-CoA reductase activity that may boost the chemosensitivity of some cancers is inhibition of the ABCB1 (a.k.a. MDR1) drug transporter;^{9, 12} over-expression of this transporter, which can extrude a range of cytotoxic agents from cancer cells, is a common cause of chemoresistance in advanced cancers.²⁶ Lovastatin's impact in this regard is only meaningful at low micromolar concentrations.

The relevance of these findings to standard clinical use of statins is dubious, as the statin dose schedules recommended for controlling LDL cholesterol typically produce statin serum levels in the low to mid nanomolar range; for example, 40-200 mg lovastatin daily is reported to produce serum concentrations of lovastatin equivalents in the 50-250 nM range.²⁷ This cautious dosing is intended to minimize risk for the myotoxicity which is the most important potential adverse effect of statin therapy; this risk is clearly dose-dependent. A number of investigations have suggested that inhibition of geranylgeranylation in muscle fibers plays a key role in the muscle pain and necrosis that occasionally is induced by statin therapy.²⁸⁻³² It is notable that few if any of the studies demonstrating potentiation of cytotoxic cancer cell killing with statins have demonstrated such an effect with nanomolar statin levels – likely because such concentrations have too modest an impact on geranylgeranylation to be useful in this regard. This accords well with the fact that statin-induced myotoxicity is only observed in a minority of patients treated with standard clinical doses.

Moreover, a recent meta-analysis of prolonged randomized controlled studies of standard statin therapy has failed to observe any impact on cancer mortality during the trials.³³ On the other hand, epidemiological evidence does suggest that survival in cancer patients receiving statins may be modestly better overall, and that outcomes in breast and prostate cancer in particular may be benefited.³⁴⁻³⁸ Even if standard statin usage does have a favorable impact on survival in some cancers, it is not clear whether chemosensitization contributes to this effect – and in any case, it is reasonable to conclude that standard clinical statin doses would be unlikely to optimize the impact of statins on chemosensitivity.

Some cell culture investigations conclude that the concentration of statin capable of markedly inhibiting cholesterol synthesis is considerably lower than the micromolar concentrations which notably inhibit geranylgeranylation – likely because geranylgeranyl transferase has a very high affinity for its substrate.^{39, 40} On the other hand, there is growing body of evidence that inhibition of isoprenylation reactions may be a key mechanism whereby clinical statin doses provide protection from vascular events, independent of their impact on lipoprotein levels.⁴¹⁻⁴³ It is not clear why these lines of research are in seeming conflict – perhaps the impact of clinical statin levels on isoprenylation in certain types of cells is more meaningful than in others. Presumably, effective inhibition of geranylgeranylation would require higher levels of statins in cell types which maintain relatively high intracellular pools of geranylgeranyl diphosphate, and/or high expression of geranylgeranyl transferase.

Practical Regimens for Statin Chemosensitization

In any case, the available cell culture data suggest that low micromolar concentrations of statins may be required to potentiate the cell-killing efficacy of cytotoxic agents. Since the pharmacokinetics of statins tend to show a linear dose-dependence, it seems likely that some sufficiently high oral intake of statins

could achieve low micromolar serum concentrations of these agents. However, it is reasonable to presume that such a high intake, if continued for prolonged periods, would be highly likely to evoke important myotoxicity. For this reason, the most clinically feasible way to achieve statin-mediated potentiation of cancer chemotherapy may be to administer high doses of statins for just several days at a time, just prior to and for two to three days following administration of a cytotoxin; the title of this essay refers to this strategy as “flash-potential” of cancer chemotherapy. As an example, when chemotherapy is administered once weekly, three times per month, it might be reasonable to give high-dose statins for three days three times monthly – for one day prior to and two days following each dose of cytotoxin. Such a regimen would likely mitigate risk for myotoxicity, as high statin levels would only be present about 30% of the time.

This “flash-potential” proposal is not novel – back in 1999, Holt and colleagues wrote that “lovastatin might therefore be used in cycles with chemotherapeutic agents to increase tumor cell kill, rather than being used over prolonged periods to inhibit tumor proliferation. This would minimize the adverse effects of high-dose lovastatin administered over prolonged periods and would increase its utility in cancer chemotherapy.”²

Fortunately, several phase I or II clinical trials of high-dose statins in cancer patients have already been conducted, in an effort to determine what dose ranges might be clinically tolerable.⁴⁴⁻⁴⁸ These investigations were ultimately intended to determine whether intermittent administration of statins *alone* could achieve clinically worthwhile cancer control – since micromolar statin doses by themselves have slowed cellular proliferation or enhanced apoptosis in some cancer cell lines. Few objective responses were noted in these investigations – in a handful of cases, temporary stasis of cancer growth was reported – likely in part because statins could only be administered intermittently at the doses tested. It thus seems to have been concluded that statin therapy *per se* was unlikely to have clinically important utility in oncology, as phase III studies have not been reported. Nonetheless, these studies established that various high-dose intermittent statin protocols could be administered with an acceptable side effect profile. In particular, lovastatin administered in the range of 25-35 mg/kg daily, in multiple divided doses, for seven consecutive days, once monthly, appears to be reasonably well tolerated, with nausea, diarrhea, myalgia and muscle weakness, typically of grade 1 or 2 intensity, as the most common side effects.⁴⁴⁻⁴⁶ In the study by Thibault et al., peak serum concentrations of active lovastatin metabolites as high as 3.9 μM were measured; in light of cell culture data, it seems likely that such concentrations would be sufficient to achieve a measure of chemosensitization in many cancers.⁴⁴ This is not unreasonable, given that the doses employed were about 10-fold higher than the highest dose of lovastatin used for cholesterol control. When administered for 21 consecutive days, every 28 days, the maximal tolerated dose of lovastatin was found to be 7.5 mg/kg.⁴⁸

Concurrent supplementation with ubiquinone was employed with some of these protocols, and in one such study the investigators had an impression that this reduced the incidence and severity of muscle symptoms;⁴⁴ however, this conclusion is hard to square with the cited evidence that suppressed geranylgeranylation may be largely responsible for statin-induced myopathy. In any case, concurrent supplementation with ubiquinone appears prudent during high-dose statin therapy, to alleviate any concern that induced ubiquinone deficiency might compromise cardiac performance or lead to other adverse effects; there is no reason to suspect that ubiquinone deficiency plays a role in the chemosensitizing impact of statins.

This author has been able to trace only two studies in which high-dose statins were administered in conjunction with chemotherapy. In a dose-finding study enrolling patients with relapsed or refractory myeloma or lymphoma, simvastatin was administered for 7 days, following which chemotherapy with VAD or CHOP was commenced.⁴⁹ The maximal tolerated dose was found to be 15 mg/kg; the dose-limiting toxicities were grade 3 gastrointestinal effects and neutropenic fever. 30% of the patients achieved a clinical response; 3 of these 7 patients had been previously judged refractory to the chemotherapy regimen they received – suggesting that simvastatin may have aided their subsequent response. However, in a follow-up study by this same group, which enrolled 12 patients with refractory or relapsed myeloma, only one of the patients achieved remission with the simvastatin-VAD regimen, and further study was discontinued.⁵⁰ It is not unlikely that the efficacy of this regimen was compromised by the fact that statin administration was discontinued before the cytotoxic agents were given; in the cell culture studies demonstrating synergism between statins and cytotoxins, these agents have been present simultaneously.

In light of these considerations, it would seem worthwhile to evaluate a flash potentiation regimen in which 30 mg/kg/day lovastatin, administered orally in divided doses, is given for three consecutive days in conjunction with chemotherapy, the lovastatin administration to commence one day prior to the chemotherapy.

In light of the nausea and diarrhea occasionally observed with high-dose statin regimens – likely reflecting privileged access of intestinal epithelium to oral statins – the possibility that high-dose statins might potentiate the gastrointestinal toxicity of concurrent chemotherapy merits consideration. Indeed, in a phase II trial evaluating continual normal-dose pravastatin (40 mg/day) as an adjuvant to a chemotherapy regimen in patients with advanced gastric carcinoma, a non-significant trend toward increased GI side effects was noted in the active pravastatin arm.⁵¹

Adjunctive Potential of Tocotrienols

Some investigators have proposed that a somewhat more tumor-selective suppression of HMG-CoA reductase activity might be achieved if statin therapy is complemented by administration of certain naturally occurring isoprenoids, such as tocotrienols.^{52, 53} Cancers, particularly those that are aggressive, tend to express high HMG-CoA reductase activity because they are resistant to sterol-mediated suppression of reductase expression;^{54, 55} this phenomenon mandates the use of high, potentially toxic statin doses if isoprenylation reactions in cancers are to be minimized. However, cancers retain sensitivity to another feedback mechanism controlling reductase activity. Farnesol and various other isoprenoid compounds serve as a feedback signal that decreases the translation of HMG-CoA reductase mRNA while also promoting accelerated degradation of the reductase protein; tocotrienols have this effect, and some evidence suggests that their action in this regard may be somewhat cancer-selective.^{52, 56, 57} Hence, it has been suggested that co-administration of a statin and tocotrienols may achieve a more potent inhibition of HMG-CoA reductase activity in tumors than in healthy cells, thereby down-regulating isoprenylation more selectively in tumors while minimizing toxic risk.^{52, 53} Whereas statin administration tends to up-regulate HMG-CoA reductase expression by relieving feedback inhibition, tocotrienols block this up-regulation in cancer cells.⁵⁸ Anti-proliferative synergism between statins and tocotrienols has indeed been reported in cancer cell lines.^{53, 58-61}

Prolonged administration of high-dose tocotrienols (which owing to their lipid solubility require a number of days to achieve equilibrium tissue levels), in conjunction with intermittent administration of high-dose statins, might ultimately be a preferable approach to flash-potential of chemotherapy if the impact of tocotrienols on HMG-CoA reductase expression is indeed tumor-selective. Inclusion of tocotrienols in the regimen could either be used to enable a dose reduction for the lovastatin – presumably lessening toxicity; to amplify the net impact of the regimen on isoprenylation within the cancer; or preferably both. And, propitiously, tocotrienols have cancer-retardant and chemosensitizing effects of their own, unlikely to be fully attributable to isoprenylation inhibition, that could well complement the chemosensitizing impact of isoprenylation suppression.⁶²⁻⁶⁷ The pharmacokinetics of high-dose tocotrienol supplementation are now being studied in humans.^{68, 69}

References

- (1) Morris TJ, Palm SL, Furcht LL, Buchwald H. Effect of lovastatin alone and as an adjuvant chemotherapeutic agent on hepatoma tissue culture-4 cell growth. *Ann Surg Oncol* 1995 May;2(3):266-74.
- (2) Agarwal B, Bhendwal S, Halmos B, Moss SF, Ramey WG, Holt PR. Lovastatin augments apoptosis induced by chemotherapeutic agents in colon cancer cells. *Clin Cancer Res* 1999 August;5(8):2223-9.
- (3) Feleszko W, Jakobisiak M. Lovastatin augments apoptosis induced by chemotherapeutic agents in colon cancer cells. *Clin Cancer Res* 2000 March;6(3):1198-9.
- (4) Feleszko W, Mlynarczuk I, Olszewska D et al. Lovastatin potentiates antitumor activity of doxorubicin in murine melanoma via an apoptosis-dependent mechanism. *Int J Cancer* 2002 July 1;100(1):111-8.
- (5) Holstein SA, Hohl RJ. Synergistic interaction of lovastatin and paclitaxel in human cancer cells. *Mol Cancer Ther* 2001 December;1(2):141-9.
- (6) Kozar K, Kaminski R, Legat M et al. Cerivastatin demonstrates enhanced antitumor activity against human breast cancer cell lines when used in combination with doxorubicin or cisplatin. *Int J Oncol* 2004 May;24(5):1149-57.
- (7) Bocci G, Fioravanti A, Orlandi P et al. Fluvastatin synergistically enhances the antiproliferative effect of gemcitabine in human pancreatic cancer MIAPaCa-2 cells. *Br J Cancer* 2005 August 8;93(3):319-30.
- (8) Calabro A, Tai J, Allen SL, Budman DR. In-vitro synergism of m-TOR inhibitors, statins, and classical chemotherapy: potential implications in acute leukemia. *Anticancer Drugs* 2008 August;19(7):705-12.

- (9) Martirosyan A, Clendening JW, Goard CA, Penn LZ. Lovastatin induces apoptosis of ovarian cancer cells and synergizes with doxorubicin: potential therapeutic relevance. *BMC Cancer* 2010;10:103.
- (10) Taylor-Harding B, Orsulic S, Karlan BY, Li AJ. Fluvastatin and cisplatin demonstrate synergistic cytotoxicity in epithelial ovarian cancer cells. *Gynecol Oncol* 2010 December;119(3):549-56.
- (11) Wang W, Le W, Cho DY, Hwang PH, Upadhyay D. Novel effects of statins in enhancing efficacy of chemotherapy in vitro in nasopharyngeal carcinoma. *Int Forum Allergy Rhinol* 2011 July;1(4):284-9.
- (12) Follet J, Corcos L, Baffet G et al. The association of statins and taxanes: an efficient combination trigger of cancer cell apoptosis. *Br J Cancer* 2012 February 14;106(4):685-92.
- (13) Chen J, Lan T, Hou J et al. Atorvastatin sensitizes human non-small cell lung carcinomas to carboplatin via suppression of AKT activation and upregulation of TIMP-1. *Int J Biochem Cell Biol* 2012 May;44(5):759-69.
- (14) Feleszko W, Mlynarczuk I, Balkowiec-Iskra EZ et al. Lovastatin potentiates antitumor activity and attenuates cardiotoxicity of doxorubicin in three tumor models in mice. *Clin Cancer Res* 2000 May;6(5):2044-52.
- (15) Ahn KS, Sethi G, Aggarwal BB. Reversal of chemoresistance and enhancement of apoptosis by statins through down-regulation of the NF-kappaB pathway. *Biochem Pharmacol* 2008 February 15;75(4):907-13.
- (16) Rozados VR, Hinrichsen LI, Binda MM et al. Lovastatin enhances the antitumoral and apoptotic activity of doxorubicin in murine tumor models. *Oncol Rep* 2008 May;19(5):1205-11.
- (17) Sadeghi-Aliabadi H, Minaiyan M, Dabestan A. Cytotoxic evaluation of doxorubicin in combination with simvastatin against human cancer cells. *Res Pharm Sci* 2010 July;5(2):127-33.
- (18) Fromigue O, Hamidouche Z, Marie PJ. Statin-induced inhibition of 3-hydroxy-3-methyl glutaryl coenzyme a reductase sensitizes human osteosarcoma cells to anticancer drugs. *J Pharmacol Exp Ther* 2008 May;325(2):595-600.
- (19) Roudier E, Mistafa O, Stenius U. Statins induce mammalian target of rapamycin (mTOR)-mediated inhibition of Akt signaling and sensitize p53-deficient cells to cytostatic drugs. *Mol Cancer Ther* 2006 November;5(11):2706-15.
- (20) Soma MR, Baetta R, De Renzis MR et al. In vivo enhanced antitumor activity of carmustine [N,N'-bis(2-chloroethyl)-N-nitrosourea] by simvastatin. *Cancer Res* 1995 February 1;55(3):597-602.
- (21) Huelsenbeck J, Henninger C, Schad A, Lackner KJ, Kaina B, Fritz G. Inhibition of Rac1 signaling by lovastatin protects against anthracycline-induced cardiac toxicity. *Cell Death Dis* 2011;2:e190.
- (22) Huelsenbeck SC, Schorr A, Roos WP et al. Rac1 Protein Signaling Is Required for DNA Damage Response Stimulated by Topoisomerase II Poisons. *J Biol Chem* 2012 November 9;287(46):38590-9.

- (23) Ghosh-Choudhury N, Mandal CC, Ghosh-Choudhury N, Ghosh CG. Simvastatin induces derepression of PTEN expression via NFkappaB to inhibit breast cancer cell growth. *Cell Signal* 2010 May;22(5):749-58.
- (24) Kochuparambil ST, Al-Husein B, Goc A, Soliman S, Somanath PR. Anticancer efficacy of simvastatin on prostate cancer cells and tumor xenografts is associated with inhibition of Akt and reduced prostate-specific antigen expression. *J Pharmacol Exp Ther* 2011 February;336(2):496-505.
- (25) Spampinato C, De MS, Sarnataro M et al. Simvastatin inhibits cancer cell growth by inducing apoptosis correlated to activation of Bax and down-regulation of BCL-2 gene expression. *Int J Oncol* 2012 April;40(4):935-41.
- (26) Takara K, Sakaeda T, Okumura K. An update on overcoming MDR1-mediated multidrug resistance in cancer chemotherapy. *Curr Pharm Des* 2006;12(3):273-86.
- (27) Lewis KA, Holstein SA, Hohl RJ. Lovastatin alters the isoprenoid biosynthetic pathway in acute myelogenous leukemia cells in vivo. *Leuk Res* 2005 May;29(5):527-33.
- (28) Flint OP, Masters BA, Gregg RE, Durham SK. HMG CoA reductase inhibitor-induced myotoxicity: pravastatin and lovastatin inhibit the geranylgeranylation of low-molecular-weight proteins in neonatal rat muscle cell culture. *Toxicol Appl Pharmacol* 1997 July;145(1):99-110.
- (29) Sakamoto K, Honda T, Yokoya S, Waguri S, Kimura J. Rab-small GTPases are involved in fluvastatin and pravastatin-induced vacuolation in rat skeletal myofibers. *FASEB J* 2007 December;21(14):4087-94.
- (30) Cao P, Hanai J, Tanksale P, Imamura S, Sukhatme VP, Lecker SH. Statin-induced muscle damage and atrogin-1 induction is the result of a geranylgeranylation defect. *FASEB J* 2009 September;23(9):2844-54.
- (31) Johnson TE, Zhang X, Bleicher KB et al. Statins induce apoptosis in rat and human myotube cultures by inhibiting protein geranylgeranylation but not ubiquinone. *Toxicol Appl Pharmacol* 2004 November 1;200(3):237-50.
- (32) Araki M, Maeda M, Motojima K. Hydrophobic statins induce autophagy and cell death in human rhabdomyosarcoma cells by depleting geranylgeranyl diphosphate. *Eur J Pharmacol* 2012 January 15;674(2-3):95-103.
- (33) Emberson JR, Kearney PM, Blackwell L et al. Lack of effect of lowering LDL cholesterol on cancer: meta-analysis of individual data from 175,000 people in 27 randomised trials of statin therapy. *PLoS ONE* 2012;7(1):e29849.
- (34) Nielsen SF, Nordestgaard BG, Bojesen SE. Statin use and reduced cancer-related mortality. *N Engl J Med* 2012 November 8;367(19):1792-802.
- (35) Kwan ML, Habel LA, Flick ED, Quesenberry CP, Caan B. Post-diagnosis statin use and breast cancer recurrence in a prospective cohort study of early stage breast cancer survivors. *Breast Cancer Res Treat* 2008 June;109(3):573-9.

- (36) Colli JL, Amling CL. High cholesterol levels are associated with reduced prostate cancer mortality rates during periods of high but not low statin use in the United States. *Urol Oncol* 2009 March;27(2):170-3.
- (37) Gutt R, Tonlaar N, Kunnavakkam R, Karrison T, Weichselbaum RR, Liauw SL. Statin use and risk of prostate cancer recurrence in men treated with radiation therapy. *J Clin Oncol* 2010 June 1;28(16):2653-9.
- (38) Marcella SW, David A, Ohman-Strickland PA, Carson J, Rhoads GG. Statin use and fatal prostate cancer: a matched case-control study. *Cancer* 2012 August 15;118(16):4046-52.
- (39) Sinensky M, Beck LA, Leonard S, Evans R. Differential inhibitory effects of lovastatin on protein isoprenylation and sterol synthesis. *J Biol Chem* 1990 November 15;265(32):19937-41.
- (40) Cho KJ, Hill MM, Chigurupati S, Du G, Parton RG, Hancock JF. Therapeutic levels of the hydroxymethylglutaryl-coenzyme A reductase inhibitor lovastatin activate ras signaling via phospholipase D2. *Mol Cell Biol* 2011 March;31(6):1110-20.
- (41) Zhou Q, Liao JK. Pleiotropic effects of statins. - Basic research and clinical perspectives -. *Circ J* 2010 May;74(5):818-26.
- (42) Hamilton PK, Hughes SM, Plumb RD et al. Statins have beneficial effects on platelet free radical activity and intracellular distribution of GTPases in hyperlipidaemia. *Clin Sci (Lond)* 2010 March;118(5):359-66.
- (43) Liu B, Zhang JY, Cao HM, Wang Q, Wang HB. Effect of rosuvastatin on ROCK activity, endothelial function, and inflammation in Asian patients with atherosclerosis. *Intern Med* 2012;51(10):1177-82.
- (44) Thibault A, Samid D, Tompkins AC et al. Phase I study of lovastatin, an inhibitor of the mevalonate pathway, in patients with cancer. *Clin Cancer Res* 1996 March;2(3):483-91.
- (45) Larner J, Jane J, Laws E, Packer R, Myers C, Shaffrey M. A phase I-II trial of lovastatin for anaplastic astrocytoma and glioblastoma multiforme. *Am J Clin Oncol* 1998 December;21(6):579-83.
- (46) Kim WS, Kim MM, Choi HJ et al. Phase II study of high-dose lovastatin in patients with advanced gastric adenocarcinoma. *Invest New Drugs* 2001;19(1):81-3.
- (47) Sleijfer S, van der Gaast A, Planting AS, Stoter G, Verweij J. The potential of statins as part of anti-cancer treatment. *Eur J Cancer* 2005 March;41(4):516-22.
- (48) Knox JJ, Siu LL, Chen E et al. A Phase I trial of prolonged administration of lovastatin in patients with recurrent or metastatic squamous cell carcinoma of the head and neck or of the cervix. *Eur J Cancer* 2005 March;41(4):523-30.
- (49) van der Spek E, Bloem AC, van de Donk NW et al. Dose-finding study of high-dose simvastatin combined with standard chemotherapy in patients with relapsed or refractory myeloma or lymphoma. *Haematologica* 2006 April;91(4):542-5.

- (50) van der Spek E, Bloem AC, Sinnige HA, Lokhorst HM. High dose simvastatin does not reverse resistance to vincristine, adriamycin, and dexamethasone (VAD) in myeloma. *Haematologica* 2007 December;92(12):e130-e131.
- (51) Konings IR, van der Gaast A, van der Wijk LJ, de Jongh FE, Eskens FA, Sleijfer S. The addition of pravastatin to chemotherapy in advanced gastric carcinoma: a randomised phase II trial. *Eur J Cancer* 2010 December;46(18):3200-4.
- (52) Mo H, Elson CE. Studies of the isoprenoid-mediated inhibition of mevalonate synthesis applied to cancer chemotherapy and chemoprevention. *Exp Biol Med (Maywood)* 2004 July;229(7):567-85.
- (53) McAnally JA, Gupta J, Sodhani S, Bravo L, Mo H. Tocotrienols potentiate lovastatin-mediated growth suppression in vitro and in vivo. *Exp Biol Med (Maywood)* 2007 April;232(4):523-31.
- (54) Elson CE, Peffley DM, Hentosh P, Mo H. Isoprenoid-mediated inhibition of mevalonate synthesis: potential application to cancer. *Proc Soc Exp Biol Med* 1999 September;221(4):294-311.
- (55) Hentosh P, Yuh SH, Elson CE, Peffley DM. Sterol-independent regulation of 3-hydroxy-3-methylglutaryl coenzyme A reductase in tumor cells. *Mol Carcinog* 2001 November;32(3):154-66.
- (56) Parker RA, Pearce BC, Clark RW, Gordon DA, Wright JJ. Tocotrienols regulate cholesterol production in mammalian cells by post-transcriptional suppression of 3-hydroxy-3-methylglutaryl-coenzyme A reductase. *J Biol Chem* 1993 May 25;268(15):11230-8.
- (57) Peffley DM, Gayen AK. Plant-derived monoterpenes suppress hamster kidney cell 3-hydroxy-3-methylglutaryl coenzyme a reductase synthesis at the post-transcriptional level. *J Nutr* 2003 January;133(1):38-44.
- (58) Yang Z, Xiao H, Jin H, Koo PT, Tsang DJ, Yang CS. Synergistic actions of atorvastatin with gamma-tocotrienol and celecoxib against human colon cancer HT29 and HCT116 cells. *Int J Cancer* 2010 February 15;126(4):852-63.
- (59) Wali VB, Sylvester PW. Synergistic antiproliferative effects of gamma-tocotrienol and statin treatment on mammary tumor cells. *Lipids* 2007 December;42(12):1113-23.
- (60) Wali VB, Bachawal SV, Sylvester PW. Combined treatment of gamma-tocotrienol with statins induce mammary tumor cell cycle arrest in G1. *Exp Biol Med (Maywood)* 2009 June;234(6):639-50.
- (61) Sylvester PW. Synergistic anticancer effects of combined gamma-tocotrienol with statin or receptor tyrosine kinase inhibitor treatment. *Genes Nutr* 2012 January;7(1):63-74.
- (62) Luk SU, Yap WN, Chiu YT et al. Gamma-tocotrienol as an effective agent in targeting prostate cancer stem cell-like population. *Int J Cancer* 2011 May 1;128(9):2182-91.
- (63) Kannappan R, Gupta SC, Kim JH, Aggarwal BB. Tocotrienols fight cancer by targeting multiple cell signaling pathways. *Genes Nutr* 2012 January;7(1):43-52.

- (64) Yang Z, Lee MJ, Zhao Y, Yang CS. Metabolism of tocotrienols in animals and synergistic inhibitory actions of tocotrienols with atorvastatin in cancer cells. *Genes Nutr* 2012 January;7(1):11-8.
- (65) Sylvester PW, Wali VB, Bachawal SV, Shirode AB, Ayoub NM, Akl MR. Tocotrienol combination therapy results in synergistic anticancer response. *Front Biosci* 2011;16:3183-95.
- (66) Ling MT, Luk SU, Al-Ejeh F, Khanna KK. Tocotrienol as a potential anticancer agent. *Carcinogenesis* 2012 February;33(2):233-9.
- (67) Husain K, Francois RA, Yamauchi T, Perez M, Sebti SM, Malafa MP. Vitamin E delta-tocotrienol augments the antitumor activity of gemcitabine and suppresses constitutive NF-kappaB activation in pancreatic cancer. *Mol Cancer Ther* 2011 December;10(12):2363-72.
- (68) Patel V, Rink C, Gordillo GM et al. Oral tocotrienols are transported to human tissues and delay the progression of the model for end-stage liver disease score in patients. *J Nutr* 2012 March;142(3):513-9.
- (69) Ling MT, Luk SU, Al-Ejeh F, Khanna KK. Tocotrienol as a potential anticancer agent. *Carcinogenesis* 2012 February;33(2):233-9.